

## Emory LBD Study PDBP Study ID 283

BIOSPECIMEN COLLECTION & PROCESSING

## Overview

- 1. Specimen uniformity and quality
- 2. Site Equipment
- 3. Procedures
  - Kit Contents and Ordering
  - Sample Labelling
  - Sample Collection and Processing
  - Shipping Samples
  - Non-Conformance
- 4. Contact Information

# Specimen Uniformity and Quality

GENERAL REMINDERS

## Specimen Standardization and Quality

#### Most biomarkers are sensitive to *time* and *temperature*

- Standardization of processing across sites is key
- Specimens must be processed within 2 hours of collection
- Reference the BioSEND Biomarker Specimen Collection, Processing, and Shipment Manual as needed
- Do not replace or supplement any kit components without first receiving approval from BioSEND/NINDS

Questions? Email biosend@iu.edu

## Site Consumables and Equipment

#### Sites will need to supply the following items:

- Gloves
- Alcohol wipes
- Butterfly needles
- Tourniquet
- Gauze pads
- Bandages
- Microcentrifuge tube rack
- Sharps bin and lid

- Crushed ice
- Pipettes and pipette tips
- Centrifuge capable of maintaining 4°C
- -80°C Freezer
- Dry ice

## Procedures

MAINTAINING SPECIMEN UNIFORMITY AND QUALITY

## Biospecimen Collection Protocol

|                            | BL | 12M | 24M | 36M | 48M |
|----------------------------|----|-----|-----|-----|-----|
| Buffy coat<br>(2 aliquots) | X  | X   | X   | X   | X   |
| Plasma<br>(6 x 1ml)        | X  | X   | X   | X   | X   |
| Serum<br>(6 x 1ml)         | X  | X   | X   | X   | X   |
| RNA<br>(2 x 2.5ml)         | X  | X   | X   | X   | X   |
| Whole Blood<br>(1 x 3ml)   | X  | X   | X   | X   | X   |
| CSF<br>(10 x 1ml)          | X  |     | X   |     | X   |

## Kit Contents and Ordering

- All sites will be sent a Supplemental Kit with their first kit shipment
  - Contains extra blood collection tubes, processing supplies, and LP needles
  - May be used to replace items in study visit kits
- Study Visit Kits should be ordered as soon as visits are planned
  - Contains collection, processing, and shipping supplies specific to each visit
  - Includes barcoded labels
  - The supplies/labels in each study visit kit are intended for that visit only

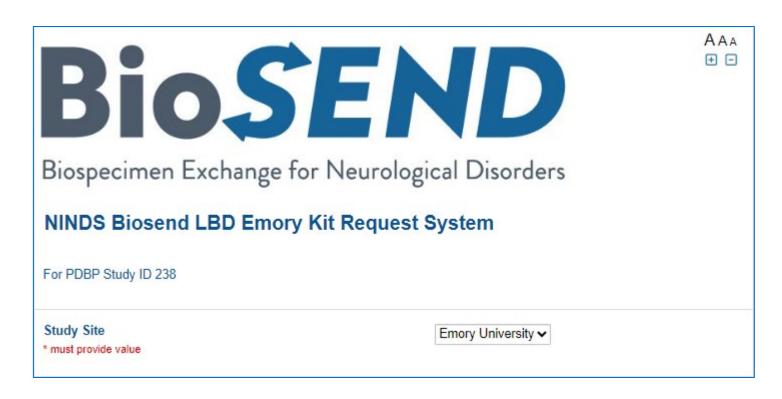
## Kit Contents and Ordering – REDCap Survey

 http://kits.iu.edu/biosend/Em oryLBD

Order kits online through the Kit Request Module for:

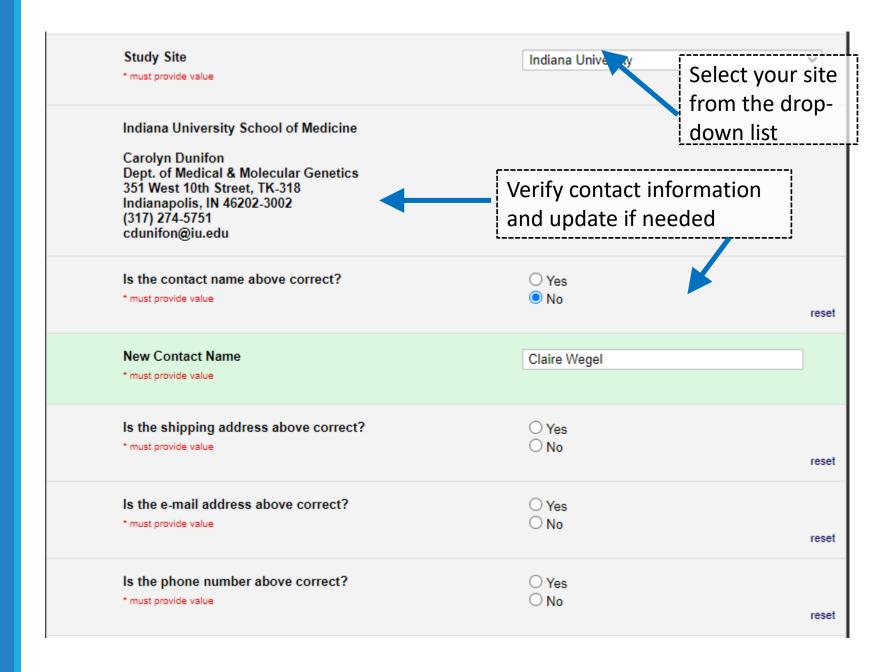
- Blood & CSF kits
- Extra Supplies

Please provide as much notice as possible when ordering kits and/or supplies.



## Kit Contents and Ordering: Confirm Site Info

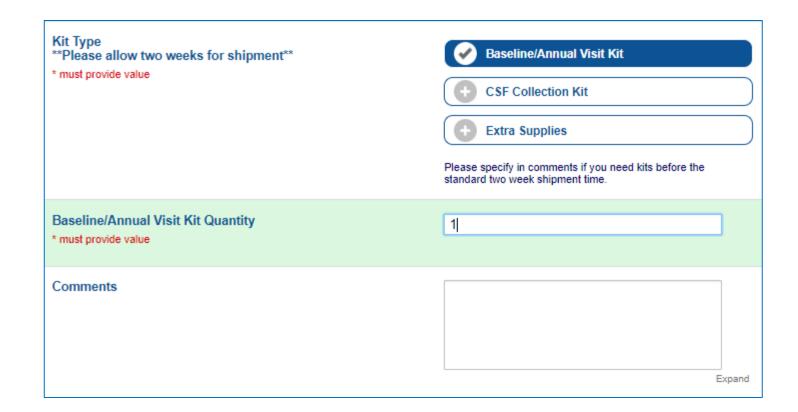
LBD Kit Request Module



## Kit Contents and Ordering: Kit Types

#### LBD Kit Request Module

- Kits are not specific to a subject or time point. After collection, sites will indicate the subject and time point to which BioSEND should link the samples.
- CSF processing kits are ordered independently of blood kits, as CSF is not required
- All specimen labels (including CSF) will be included in the Blood Collection Kit. If CSF is not collected at a visit, you may discard these extra labels



# Kit Contents and Ordering: Kit Breakdown

LBD Kit Request Module

#### Comments Expand Baseline/Annual Visit Collection Kit 15 - Siliconized cryogenic vial, 2ml Kit contents of selected 2 - Serum (glass) tube, 10ml 2 - EDTA (glass) tube, 10ml kit will appear at the 1 - EDTA (plastic) tube, 3ml bottom of the page 2 - PAXGene® tube, 2.5ml 7 - Bubble-tube sleeve 2 - Disposable pipet, 3ml 1 - Cryobox, 25 cell 2 - Biohazard bag w/ absorbent sheet 1 - Fragile label 1 - UN3373 label

1 - Dry ice label

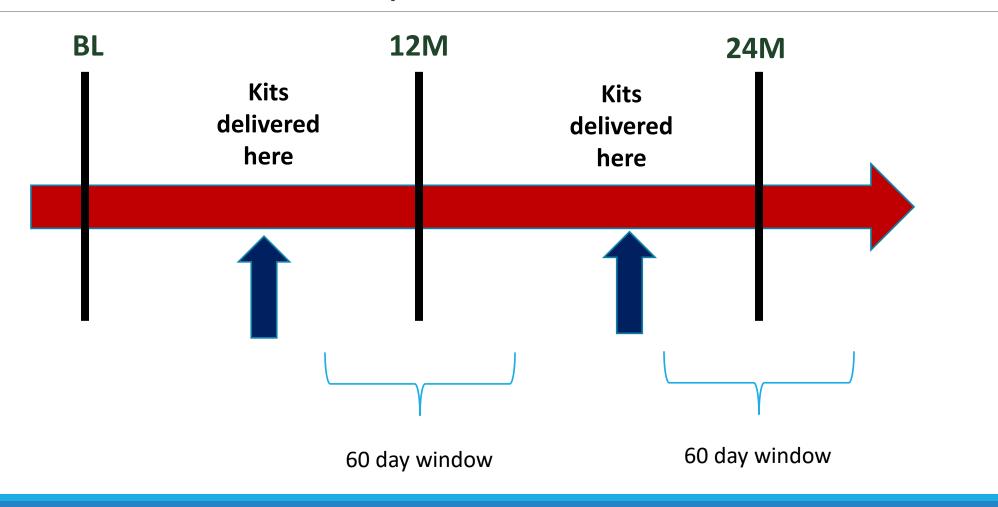
1 - Airway bill envelope1 - Frozen shipper

1 - Label set (kit & specimen labels)

## Automatic Kit Shipments

- After subject completes baseline visit and BioSEND receives BL samples, BioSEND sets up automated kit sending schedule for subject's subsequent visits
- Schedule gives 2 month window around the longitudinal study visit target (1 month on either side)
- BioSEND will send kits prior to start of study window
  - Reduces effort for study coordinators
  - Sites only need to order kits if visit will occur AHEAD of the study visit window
- All study visit target dates are determined from Baseline Visit (not from last study visit date)

## Automatic Kit Shipments

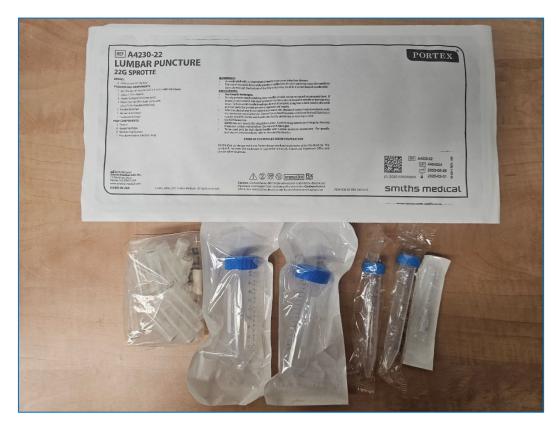


## Kit Contents and Ordering: Blood Kits



## Kit Contents and Ordering: CSF Kits

CSF: LP Tray:





## Collection Volumes

Total blood and CSF volumes

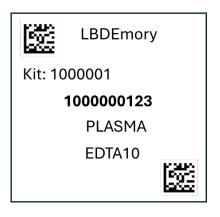
| Sample Type                           | Amount |
|---------------------------------------|--------|
| Whole Blood for RNA                   | 5 ml   |
| Whole Blood for Plasma and Buffy Coat | 20 ml  |
| Whole Blood for Serum                 | 20 ml  |
| Whole Blood for Banking               | 3 ml   |
| Cerebrospinal Fluid                   | 10 ml  |

## Sample Labelling: Example Labels

#### Labels are provided by Indiana University

- Please check that all samples are properly labelled to ensure correct identification by IU
- If do not have enough labels to complete a visit, please contact IU immediately
- Labelling the tubes during processing prevents sample mix-ups





## Sample Labelling: Label Placement

#### Please...

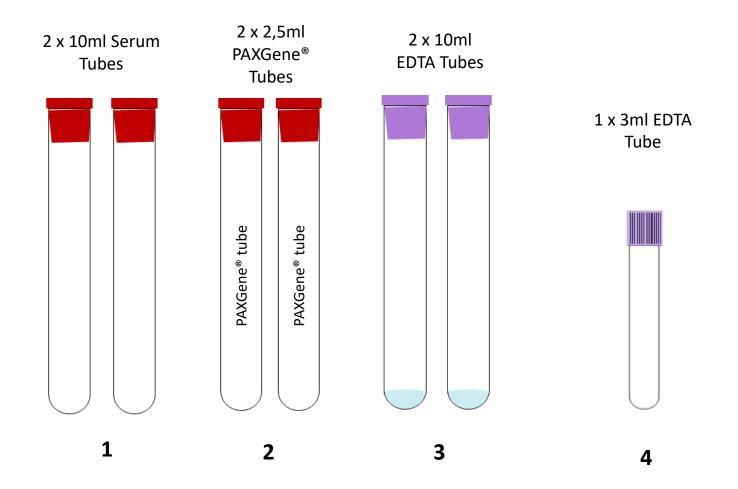
- Label all collection and aliquot tubes before cooling, collecting, processing, or freezing samples
- Label only 1 subject's tubes at a time to avoid mix-ups
- Wrap the label around the tube horizontally label position is important for all tube types
- Make sure the label is completely adhered by rolling between your fingers



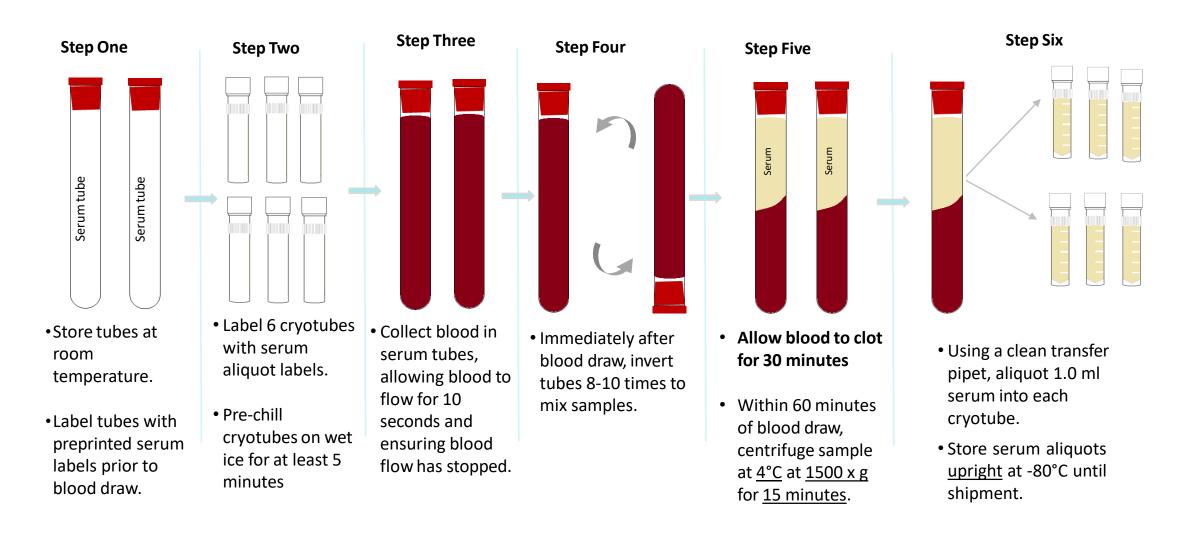


## Sample Collection and Processing

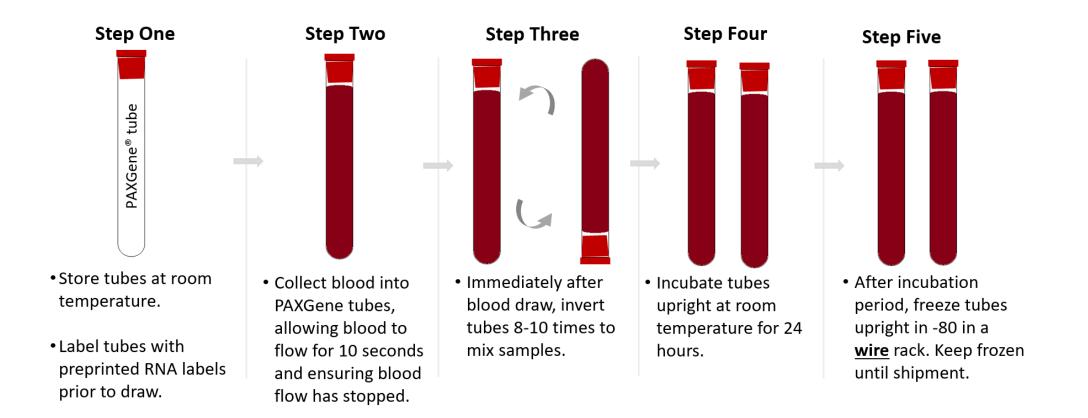
**Blood Tube Draw Order** 



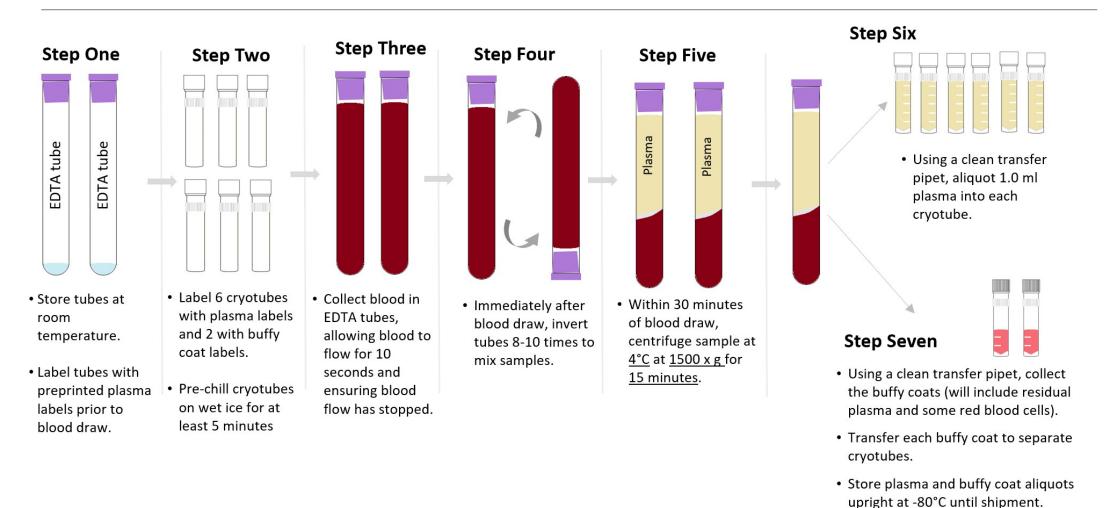
## Sample Collection and Processing: Serum



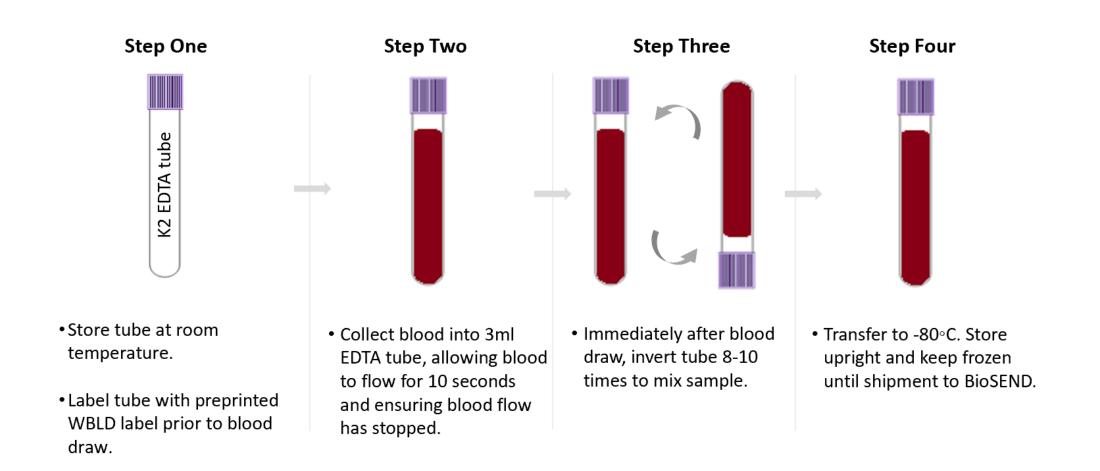
## Sample Collection and Processing: Whole blood RNA



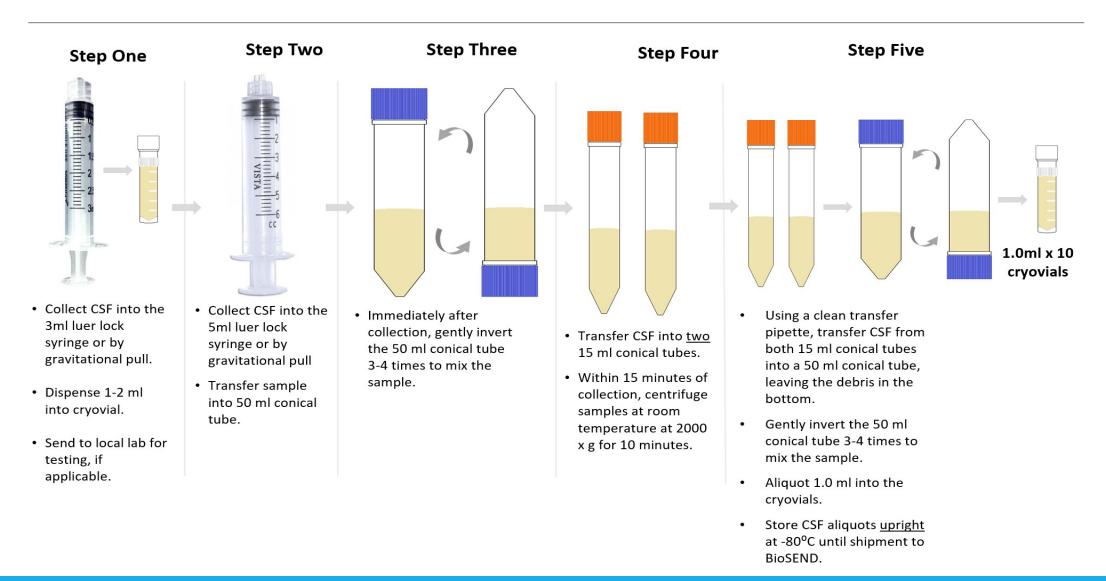
### Sample Collection and Processing: Plasma & Buffy Coat



### Sample Collection and Processing: Whole Blood



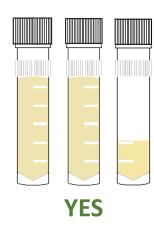
## Sample Collection and Processing: CSF



## Sample Collection and Processing: Aliquots

Filling biomarker serum, plasma, and CSF aliquots:

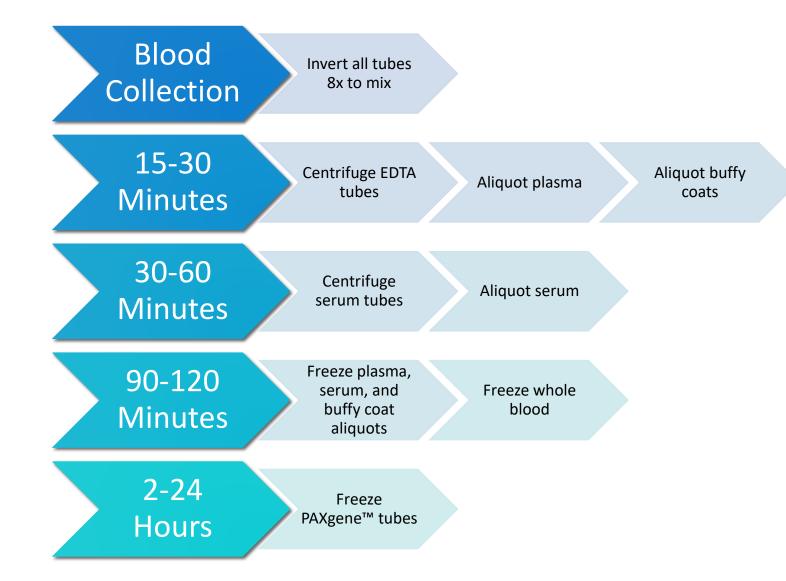
- Fill as many cryovials as possible to 1.0 ml (plasma, serum & CSF)
- Over-filled vials may burst in freezer!
- Ship ALL material to IU, even if final vial is less than standard volume





# Sample Collection and Processing: Timeline

Timeline for blood processing



## Sample Collection and Processing: Issue #1

#### **Troubleshooting Blood Collection**

#### Issue #1: Tube with little/no vacuum

- Always check expiration date on the tube before beginning blood draw and discard expired tubes
- Store tubes at "room temperature" extreme temperature can affect vacuum
- Keep extra vacutainer tubes from supplemental kit nearby during blood draw to replace "bad" tubes
- If this is a frequent occurrence, report tube type and lot number to IU.

## Sample Collection and Processing: Issue #2

#### **Troubleshooting Blood Collection**

Issue #2: Hemolyzed serum and/or plasma caused by incorrect collection

| Cause: Blood Collection Methods   | Corrective Action  |  |  |
|---|--|--|--|
| Improper venipuncture site  | Draw from median cubital, basalic, and cephalic veins from antecubital region of arm   |  |  |
| Prolonged tourniquet use  | Tourniquet should be released after no more than 1 min, excessive fist clenching should be avoided   |  |  |
| Not allowing alcohol to dry on skin before venipuncture                       | Without touching, allow the venipuncture site to air dry   |  |  |
| Use of too large/small bore needle resulting in excess force applied to blood | Avoid using too small/large needle. Needle size dependent on the subject's physical characteristics & amount of blood to be drawn. Most commonly used sizes are 19 – 23. |  |  |
| Pulling/pushing plunger too fast while drawing/transferring blood             | Avoid drawing the syringe plunger too forcefully when collecting blood   |  |  |
| Ensure all blood collection assemblies are fitted securely, to avoid frothing |  |  |  |

For more information, visit: http://www.bd.com/vacutainer/pdfs/techtalk/TechTalk\_Jan2004\_VS7167.pdf

### Sample Collection and Processing: Issue #2 continued

#### **Troubleshooting Blood Collection**

Issue #2: Hemolyzed serum and/or plasma caused by incorrect processing

| Cause: Sample Processing Methods                | Corrective Actions  |  |
|---|---|--|
| Vigorous mixing/shaking                         | Gently invert blood collection tube when mixing additive with specimen, follow guidelines in Biologics Manual regarding number of times to invert each type of tube |  |
| Not allowing serum to clot for recommended time | Serum tubes without clot activator should be allowed to clot for 60 min in a vertical position  |  |
| Exposure to excessive heat or cold              | Keep samples at ambient temperature until processing  |  |
| Prolonged contact of serum/plasma with cells    | Do not store uncentrifuged samples beyond recommended time  |  |

For more information, visit: http://www.bd.com/vacutainer/pdfs/techtalk/TechTalk\_Jan2004\_VS7167.pdf

# Sample Collection and Processing Form

Direct link:

http://kits.iu.edu/biosend/EmoryLBDSampleForm

First part captures basic subject and visit information

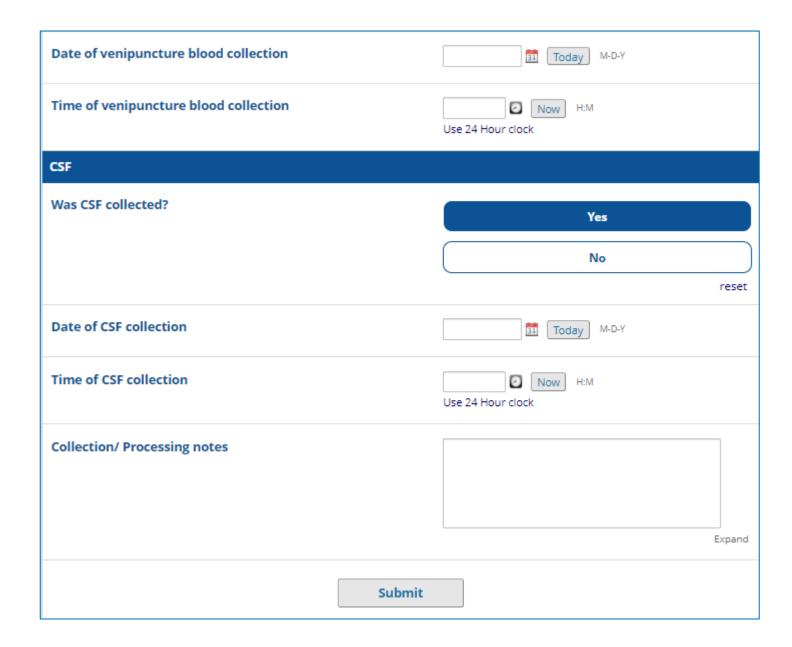


# Sample Collection and Processing Form

Direct link:

http://kits.iu.edu/biosend/EmoryLBDSampleForm

Second part captures collection information



# Sample Collection and Processing Form

Direct link:

http://kits.iu.edu/biosend/EmoryLBDSampleForm

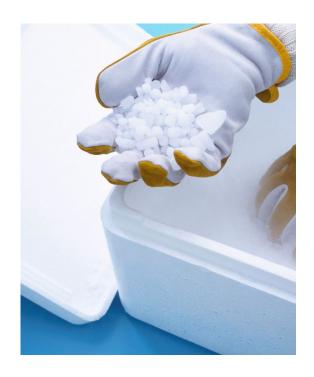
PDF form of responses will be emailed to you. Print a copy of the Frozen
Shipping Manifest and include with shipment.

| LBD Emory Frozen Shipping   | Manifest Page 6                          |
|---|--|
| Please verify/update the information below. When you clic<br>Shipping Manifest will be emailed to you for Subject [subj |  |
| Please print a copy of that document and include it in the  | shipping container.                      |
| Study Site:   | Emory University                         |
| GUID:   |  |
| Visit:  | ○ BL<br>○ 12M<br>○ 24M<br>○ 36M<br>○ 48M |
| Kit number:   |  |
| Date of blood collection:   |  |
| Date of CSF collection:   |  |
| SERUM   |  |
| Number of SERUM aliquots shipped:   |  |
| RNA   |  |
| Number of PAXGene™ tubes shipped:   |  |
| PLASMA EDTA   |  |
| Number of PLASMA EDTA aliquots shipped:   |  |
| Number of BUFFY COAT aliquots shipped:  |  |
|   |  |

## Shipping Frozen Samples: Tips

#### Packing and Shipping Frozen Samples

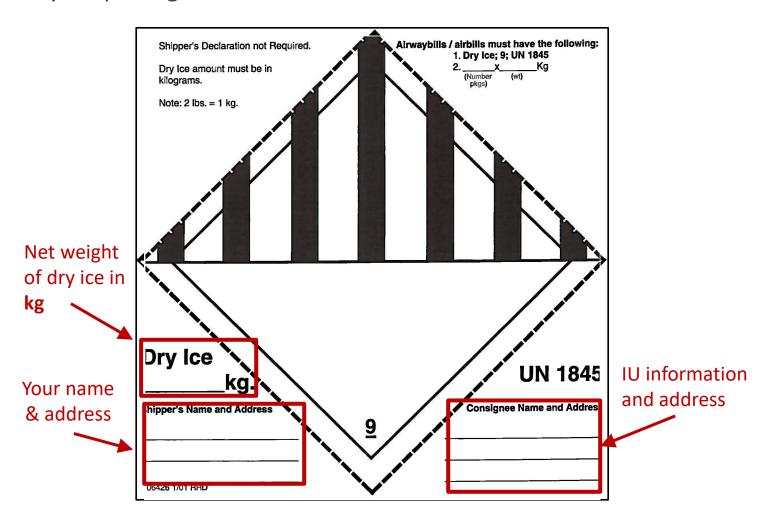
- Serum, plasma, buffy coats, CSF, whole blood and RNA all ship frozen
- Ship frozen samples on dry ice
- Frozen samples should be shipped only Monday through Wednesday
- Always fill carton to top with dry ice
- Do not pack shipment until the day of pickup



## Shipping Samples

Packing and Shipping Frozen Samples

Class 9 Dry Ice Label should not be covered with other stickers and must be completed, or UPS will reject/return your package!



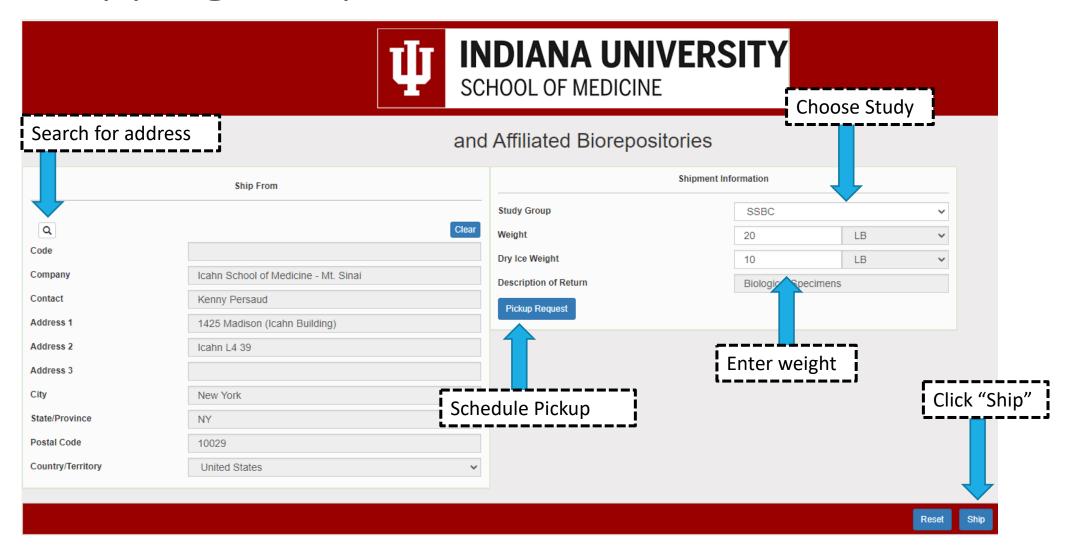
## Shipping Samples: Frozen

Packing and Shipping Frozen Samples





## Shipping Samples – UPS: <a href="https://kits.iu.edu/UPS">https://kits.iu.edu/UPS</a>



## Shipping Samples via UPS

**IU UPS ShipExec Shipping Portal** 

- Print out UPS air waybill
- Ensure all elements
   (barcode, return address, etc.) printed clearly
- Fold and insert UPS air waybill into clear plastic sleeve on package

JOHN SMITH INDIANA UNIVERSITY 410 WEST 10TH STREET INDIANAPOLIS IN 46202 2 LBS

1 OF 1

RS

SHIP TO:

SCHOOL OF MEDICINE 317-278-2694 INDIANA UNIVERSITY TK 217 351 W 10TH ST

**INDIANAPOLIS IN 46202** 



IN 461 9-01



**UPS NEXT DAY AIR** 

TRACKING #: 1Z 976 R8W 84 3985 8595

1



BILLING:

DESC: Biological Specimens RETURN SERVICE

Reference No.1: 4087277

OL 20.03.09 NV45 83.0A 12/



### Non-Conformance

Non-conformance to standard procedures may reduce the utility of the biospecimens:

- Not processing serum/plasma within 2 hours of collection allows for breakdown of certain proteins and small molecules
- Over/under centrifuging changes plasma, serum, CSF composition



## Non-Conformance Reporting con't

#### Most common non-conformance issues:

- Shipment notification not sent
- Samples shipped for weekend/holiday delivery
- Sample form incomplete/inaccurate
- Low volume
- Unlabeled or mislabeled tube(s)
- Sample hemolysis



### BioSEND.org

#### On the website, you can:

- Access your study's kit request module and sample submission form
- Download the most recent version of the Manual of Procedures
- View a recording of this training
- Find information about holiday closures
- Access shipping resources

#### **Study Resources**

#### KIT REQUEST MODULE

Please follow the below link to access the Kit Request Module. This link will direct you to a REDCap database where study coordinators and staff may request kits, individual supplies, and/or labels. Please allow a total of two weeks for kit requests to be fulfilled.

Kit Request System →

#### SPECIMEN COLLECTION AND PROCESSING FORM

Please use the below link to access the collection and processing form for this protocol. This form must be completed prior to shipment. We also ask that all shipments include a physical copy of the shipping manifest portion of the form.

Specimen Collection and Processing Form →

#### MANUAL OF PROCEDURES

The below downloadable manual was created specifically for the DxCTEII study. Please feel free to explore the manual through the hyperlinked 'Table of Contents'. Questions concerning any part of the manual may be directed to biosend@iu.edu for further clarification.

Manual of Procedures &

#### TRAINING SLIDES

These slides correspond to the BioSEND DxCTEII protocol training. Training is available upon request by contacting biosend@iu.edu.

Training Slides &

#### SAMPLE SHIPPING

BioSEND can receive samples Monday-Friday, excluding holidays. Frozen samples should be shipped M-W. Ambient samples may be shipped on Th.

Generate UPS airbill or schedule pickup →
Check holiday closures →
What do I do for Friday blood draws →

### Contacts

#### **Indiana University**

General Questions/Shipment Notifications:

biosend@iu.edu

317-278-6158

Request kits:

http://kits.iu.edu/biosend/EmoryLBD